

学校编码：10384

学号：21720090153534

廈門大學

博 士 学 位 论 文

Smurf1介导的RhoB泛素化降解在DNA损伤引起的细胞凋亡过程中的作用

Role of Smurf1-mediated RhoB degradation in DNA damage-induced apoptosis

郭磊

指导教师：王洪睿

专业名称：动物学

答辩日期：2014年8月

## 厦门大学学位论文原创性声明

本人呈交的学位论文是本人在导师指导下，独立完成的研究成果。本人在论文写作中参考其他个人或集体已经发表的研究成果，均在文中以适当方式明确标明，并符合法律规范和《厦门大学研究生学术活动规范(试行)》。

另外，该学位论文为( )课题(组)的研究成果，获得( )课题(组)经费或实验室的资助，在( )实验室完成。(请在以上括号内填写课题或课题组负责人或实验室名称，未有此项声明内容的，可以不作特别声明。)

声明人(签名)：

年 月 日

# 厦门大学学位论文著作权使用声明

本人同意厦门大学根据《中华人民共和国学位条例暂行实施办法》等规定保留和使用此学位论文，并向主管部门或其指定机构送交学位论文(包括纸质版和电子版)，允许学位论文进入厦门大学图书馆及其数据库被查阅、借阅。本人同意厦门大学将学位论文加入全国博士、硕士学位论文共建单位数据库进行检索，将学位论文的标题和摘要汇编出版，采用影印、缩印或者其它方式合理复制学位论文。

本学位论文属于：

(        )1. 经厦门大学保密委员会审查核定的保密学位论文，于  
年 月 日解密，解密后适用上述授权。

(        )2. 不保密，适用上述授权。

(请在以上相应括号内打“√”或填上相应内容。保密学位论文应是已经厦门大学保密委员会审定过的学位论文，未经厦门大学保密委员会审定的学位论文均为公开学位论文。此声明栏不填写的，默认为公开学位论文，均适用上述授权。)

声明人(签名)：

年    月    日

## 摘 要

Rho家族小分子GTP酶做为细胞内许多信号转导通路中重要的分子开关，在调节细胞骨架、细胞极性、细胞运动、基因表达、细胞生长分化、膜泡运输、以及细胞周期等细胞生物学过程中发挥着重要作用。RhoA、RhoB和RhoC是Rho家族中的一个重要的亚家族，它们在氨基酸序列上具有很高的同源性并且都参与细胞骨架的重塑过程。然而，和RhoA、RhoC不同的是，RhoB可以做为抑癌基因参与应答由DNA损伤引起的细胞凋亡过程。在紫外线照射或抗癌药物等引起的DNA损伤的情况下，RhoB做为早期应答基因能够迅速表达并介导凋亡过程的发生。RhoB基因缺失小鼠的胚胎成纤维细胞（MEF）经放射线照射或抗癌药物阿霉素处理后，其细胞凋亡的比率明显低于野生型对照组。过表达RhoB的细胞具有生长缓慢的特性并更易于发生细胞凋亡。临床样本也显示随着肿瘤恶性程度的增加，RhoB蛋白的表达水平随之下降，提示RhoB介导的凋亡过程在癌症发生中可能发挥总要作用。然而，RhoB介导细胞凋亡的分子机理以及RhoB本身水平的调控的分子机制还很不清楚。

经典的Rho蛋白活性的调节方式是通过与GTP或GDP两种不同形式的鸟嘌呤核苷酸来调控的。然而，最近的研究结果表明一些Rho家族的成员也可以通过泛素化修饰来影响其蛋白水平与生物学活性。在本文的研究中，我们发现，与泛素化调节RhoA相似，C2-WW-HECT家族E3泛素连接酶中的Smurf1也可通过泛素化修饰来调控RhoB的蛋白水平，然而却对RhoC没有影响。有趣的是，与泛素化降解RhoA不同，Smurf1泛素化降解RhoB对细胞骨架的重塑过程没有明显的作用，却对细胞应答紫外线照射和化学试剂诱导的DNA损伤所引起细胞凋亡起着至关重要的作用。在DNA损伤的情况下，Smurf1通过增强自身的降解，使得RhoB的蛋白水平显著增加，进而引起细胞凋亡。Smurf1介导的RhoB泛素化降解对于细胞应答DNA损伤具有重要作用：在敲低内源Smurf1的情况下，细胞对于DNA损伤的敏感性大大增加，而在过量表达Smurf1的情况下，细胞对于DNA损伤的耐受程度明显增加，显著地抑制了DNA损伤所诱导的细胞凋亡。因此，本文发现了一个通过E3泛素连接酶Smurf1调控RhoB蛋白稳定性从而调节细胞应答DNA损伤所引起的凋亡的分子机制，为进一步深入了解与RhoB相关的肿瘤发生的分子机理提供了新的思路，并为以RhoB为分子靶点的可能的肿瘤治疗提供

了理论基础和实验依据

**关键词：**E3泛素连接酶；Smurf1；RhoB；DNA损伤；凋亡

厦门大学博硕士学位论文摘要库

## Abstract

As molecular switches, Rho family small GTPases play key roles in many signal transduction pathways that regulate cytoskeleton, cell polarity, cell migration, gene expression, cell growth and differentiation, vesicular transport, and cell cycle. RhoA, RhoB and RhoC, are three close family members that belong to the Rho family and participate in cytoskeleton remodeling. Although they share high homology in amino acid sequence, RhoB has different function from RhoA and RhoC. It has been reported that RhoB may take part in DNA damage-induced apoptosis as a tumor suppressor gene. In the event of DNA damage caused by UV or anti-cancer drugs, RhoB expression is quickly up-regulated, which is important for apoptosis process. Mouse Embryonic Fibroblast cell (MEF) isolated from RhoB-null mouse had lower apoptotic rate than control groups after UV irradiation or doxorubicin treatment. Cells ectopically expressing RhoB exhibit growth retard and are apt to apoptosis. Clinical samples also demonstrated that correlated with the malignancy of tumors, RhoB protein levels were significantly down-regulated, indicating that RhoB-mediated apoptosis may play important roles in cancer development. However, the molecular mechanism underlying is still obscure.

Classical regulation of Rho proteins is dependent on their binding status with GDP or GTP. Recent reports demonstrated that ubiquitin modification could also affect protein levels of Rho GTPase and their biological functions. In our research, we found that, similar with RhoA, RhoB but not RhoC could also be ubiquitinated by Smurf1, a member of C2-WW-HECT E3 family ubiquitin ligase. In contrast to Smurf1-mediated degradation of RhoA, degradation of RhoB by Smurf1 have no apparent effect on the remodeling of cytoskeleton. Interestingly, Smurf1-dependent degradation of RhoB is essential for DNA damage-induced apoptosis. During treatment of UV or chemical reagents, Smurf1 is down-regulated, which

subsequently up-regulated protein levels of RhoB that is important for cell apoptosis. Cells were much more susceptible to DNA damage when endogenous Smurf1 was knocked-down. When Smurf1 was overexpressed, cells were substantially more resistant to DNA damage. Thus, we identified a novel mechanism that RhoB degradation by Smurf1 determines the susceptibility of cells to DNA damage: In addition, our discovery pointed a road for further research on the molecular mechanism of RhoB-involved cancer development and potential cancer therapy way using RhoB as a target.

**Keywords:** E3 ubiquitin ligase Smurf1 RhoB DNA damage apoptosis

## 参考资料

- [1] C.M. Pickart, M.J. Eddins, Ubiquitin: structures, functions, mechanisms, *Biochim Biophys Acta* 1695(1-3) (2004) 55-72.
- [2] A. Herskho, A. Ciechanover, The ubiquitin system, *Annu Rev Biochem* 67 (1998) 425-479.
- [3] J. Smalle, R.D. Vierstra, The ubiquitin 26S proteasome proteolytic pathway, *Annu Rev Plant Biol* 55 (2004) 555-590.
- [4] R.L. Welchman, C. Gordon, R.J. Mayer, Ubiquitin and ubiquitin-like proteins as multifunctional signals, *Nat Rev Mol Cell Biol* 6(8) (2005) 599-609.
- [5] R. Groisman, J. Polanowska, I. Kuraoka, J. Sawada, M. Saijo, R. Drapkin, A.F. Kisselev, K. Tanaka, Y. Nakatani, The ubiquitin ligase activity in the DDB2 and CSA complexes is differentially regulated by the COP9 signalosome in response to DNA damage, *Cell* 113(3) (2003) 357-367.
- [6] F. Bernassola, M. Karin, A. Ciechanover, G. Melino, The HECT family of E3 ubiquitin ligases: multiple players in cancer development, *Cancer Cell* 14(1) (2008) 10-21.
- [7] C.M. Pickart, D. Fushman, Polyubiquitin chains: Polymeric protein signals, *Current Opinion in Chemical Biology* 8(6) (2004) 610 – 616.
- [8] A. Ciechanover, A.L. Schwartz, The ubiquitin system: pathogenesis of human diseases and drug targeting, *Biochim Biophys Acta* 1695(1-3) (2004) 3-17.
- [9] D. Nandi, P. Tahiliani, A. Kumar, D. Chandu, The ubiquitin-proteasome system, *J Biosci* 31(1) (2006) 137-155.
- [10] L. Hicke, R. Dunn, Regulation of membrane protein transport by ubiquitin and ubiquitin-binding proteins, *Annual Review of Cell and Developmental Biology* 19 (2003) 141 – 172.
- [11] P.J. Plant, H. Yeger, O. Staub, P. Howard, D. Rotin, The C2 domain of the ubiquitin protein ligase Nedd4 mediates  $\text{Ca}^{2+}$ -dependent plasma membrane localization, *Journal of Biological Chemistry* 272(1997) 32329 – 32336.
- [12] P.J. Lu, X.Z. Zhou, M. Shen, K.P. Lu, Function of WW domains as phosphoserine- or phosphothreonine-binding modules, *Science* 283(1999) 1325 – 1328.
- [13] C. Chen, L.E. Matesic, The Nedd4-like family of E3 ubiquitin ligases and cancer, *Cancer Metastasis Rev* 26 (2007) 587 – 604.
- [14] O. Staub, S. Dho, P. Henry, J. Correa, T. Ishikawa, J. McGlade, D. Rotin, WW domains of Nedd4 bind to the proline-rich PY motifs in the epithelial  $\text{Na}^{+}$  channel deleted in Liddle's syndrome, *EMBO J* 15(10) (1996) 2371 – 2380.
- [15] X. Lin, M. Liang, X.H. Feng, Smurf2 is a ubiquitin E3 ligase mediating proteasome-dependent degradation of Smad2 in transforming growth factor- $\beta$  signaling, *Journal of Biological Chemistry* 275(2000) 36818 – 36822.
- [16] A. Komuro, T. Imamura, M. Saitoh, Y. Yoshida, T. Yamori, K. Miyazono, et al, Negative regulation of transforming growth factor- $\beta$  (TGF- $\beta$ ) signaling by WW domain-containing protein 1 (WWP1), *Oncogene* 23(2004) 6914 – 6923.
- [17] A. Laine, Z. Ronai, Regulation of p53 localization and transcription by the HECT domain E3 ligase WWP1, *Oncogene* 26(10) (2007) 1477 – 1483.
- [18] L.C. Trotman, X. Wang, A. Alimonti, Z. Chen, J. Teruya-Feldstein, H. Yang, et al, Ubiquitination regulates PTEN nuclear import and tumor suppression, *Cell* 128 (2007) 141 – 156.
- [19] O. Micheau, S. Lens, O. Gaide, K. Alevizopoulos, J. Tschopp, NF- $\kappa$ B signals induce the expression of c-FLIP, *Molecular and Cellular Biology* 21(2001) 5299 – 5305.
- [20] H.R. Wang, Y. Zhang, B. Ozdamar, A. A. Ogunjimi, E. Alexandrova, G.H. Thomsen, et al, Regulation of cell polarity and protrusion formation by targeting RhoA for degradation, *Science* 302 (2003) 1775 – 1779.
- [21] A. Hall, The cellular functions of small GTP-binding proteins, *Science* 249(4969) (1990) 635-640.
- [22] A. Valencia, P. Chardin, A. Wittinghofer, C. Sander, The ras protein family: evolutionary tree and role of



conserved amino acids, *Biochemistry* 30(19) (1991) 4637-4648.

[23] H.R. Bourne, D.A. Sanders, F. McCormick, The GTPase superfamily: conserved structure and molecular mechanism, *Nature* 349 (1991) 117-127.

[24] M. Paduch, F. Jelen, J. Otlewski, Structure of small G proteins and their regulators, *Acta Biochim Pol* 48(4) (2001) 829-850.

[25] Y. Takai, T. Sasaki, T. Matozaki, Small GTP-binding proteins, *Physiol Rev* 81(1) (2001) 153-208.

[26] D. Vigil, J. Cherfils, K.L. Rossman, C.J. Der, Ras superfamily GEFs and GAPs: validated and tractable targets for cancer therapy, *Nature Rev Cancer* 10(12) 842-857.

[27] T. Sasaki, Y. Takai, The Rho small G protein family-Rho GDI system as a temporal and spatial determinant for cytoskeletal control, *Biochem Biophys Res Commun* 245(3) (1998) 641-645.

[28] G.A. Repasky, E.J. Chenette, C.J. Der, Renewing the conspiracy theory debate: does Raf function alone to mediate Ras oncogenesis, *Trends Cell Biol* 14(11) (2004) 639-647.

[29] J. Colicelli, Human RAS superfamily proteins and related GTPases, *Science Signaling* 250 (2004)

[30] J.H. Overmeyer, W.A. Maltese, Death pathways triggered by activated Ras in cancer cells, *Front Biosci* 16 (2011) 1693-1713.

[31] S. E. Manneville, A. Hall, Rho GTPases in cell biology, *Nature* 420 (2002) 629-635.

[32] A.J. Ridley, Rho family proteins: coordinating cell responses, *Trends Cell Biol*, 11 (2001) 471-477.

[33] O.A. Coso, M. Chiariello, J.C. Yu, H. Teramoto, P. Crespo, N. Xu, T. Miki, J.S. Gutkind, The small GTP-binding proteins Rac1 and Cdc42 regulate the activity of the JNK/SAPK signaling pathway, *Cell* 81 (1995) 1137 – 1146.

[34] A. Minden, A. Lin, F.X. Claret, A. Abo, M. Karin, Selective activation of the JNK signaling cascade and c-Jun transcriptional activity by the small GTPases Rac and Cdc42Hs, *Cell* 81 (1995) 1147 – 1157,

[35] J.B. Pereira-Leal, M.C. Seabra, Evolution of the Rab family of small GTP-binding proteins, *J Mol Biol* 313(4) (2001) 889-901.

[36] I. Jordens, M. Marsman, C. Kuijl, J. Neefjes, Rab Proteins, Connecting Transport and Vesicle Fusion, *Traffic* 6(12) (2005) 1070-1077.

[37] M. Zerial, H. McBride, Rab proteins as membrane organizers, *Nat Rev Mol Cell Biol* 2(2) (2001) 107-117.

[38] Z. Nie, D.S. Hirsch, P.A. Randazzo, Arf and its many interactors, *Curr Opin Cell Biol* 15 (2003) 396-404.

[39] J. Joseph, Ran at a glance, *J Cell Sci* 119(17) (2006) 3481-3484.

[40] H.Y. Li, K. Cao, Y. Zheng, Ran in the spindle checkpoint: a new function for a versatile GTPase, *Trends Cell Biol* 13(2003) 553-557.

[41] R.A. Kahn, J. Cherfils, M. Elias, R.C. Lovering, S. Munro, A. Schurmann, Nomenclature for the human Arf family of GTP-binding proteins: ARF, ARL, and SAR proteins, *J Cell Biol* 172 (2006) 645 – 650.

[42] D.S. Schorey, C. Chavrier, P. Chavrier, ARF proteins: roles in membrane traffic and beyond, *Nat Rev Mol Cell Biol* 7 (2006) 347 – 358.

[43] A.P. Wheeler, A.J. Ridley, Why three Rho proteins? RhoA, RhoB, RhoC, and cell motility, *Exp Cell Res* 301(1) (2004) 43-49.

[44] J. Huelsenbeck, S.C. Dreger, R. Gerhard, G. Fritz, I. Just, H. Genth, Upregulation of the immediate early gene product RhoB by exoenzyme C3 from *Clostridium limosum* and toxin B from *Clostridium difficile*, *Biochemistry* 46(16) (2007) 4923-4931.

[45] M. Huang, U. Kamasani, G.C. Prendergast, RhoB facilitates c-Myc turnover by supporting efficient nuclear accumulation of GSK-3, *Oncogene* 25(9) (2006) 1281-1289.

[46] D. Shook, R. Keller, Mechanisms, mechanics and function of epithelial-mesenchymal transitions in early development, *Mech Dev* 120(11) (2003) 1351-1383.

[47] D. Perez-Sala, P. Boya, I. Ramos, M. Herrera, K. Stamatakis, The C-terminal sequence of RhoB directs protein degradation through an endo-lysosomal pathway, *PLoS One* 4(12) (2009) e8117.

[48] B. Ozdamar, R. Bose, M. Barrios-Rodiles, H.R. Wang, Y. Zhang, J.L. Wrana, Regulation of the polarity protein Par6 by TGF $\beta$  receptors controls epithelial cell plasticity, *Science* 307(5715) (2005) 1603-1609.

- [49] P.L. Rodriguez, S. Sahay, O.O. Olabisi, I.P. Whitehead, ROCK I-mediated activation of NF-kappaB by RhoB, *Cell Signal* 19(11) (2007) 2361-2369.
- [50] J. Mazieres, V. Tillement, C. Allal, C. Clanet, L. Bobin, Z. Chen, S.M. Sebt, G. Favre, A. Pradines, Geranylgeranylated, but not farnesylated, RhoB suppresses Ras transformation of NIH-3T3 cells, *Exp Cell Res* 304(2) (2005) 354-364.
- [51] R. Karlsson, E.D. Pedersen, Z. Wang, C. Brakebusch, Rho GTPase function in tumorigenesis, *Biochim Biophys Acta* 1796(2) (2009) 91-98.
- [52] K. Asanuma, E. Yanagida-Asanuma, C. Faul, Y. Tomino, K. Kim, P. Mundel, Synaptopodin orchestrates actin organization and cell motility via regulation of RhoA signalling, *Nat Cell Biol* 8(5) (2006) 485-491.
- [53] A.P. Wheeler, A.J. Ridley, RhoB affects macrophage adhesion, integrin expression and migration, *Exp Cell Res* 313(16) (2007) 3505-3516.
- [54] M. Yoneda, Y.S. Hirokawa, A. Ohashi, K. Uchida, D. Kami, M. Watanabe, T. Yokoi, T. Shiraishi, S. Wakusawa, RhoB enhances migration and MMP1 expression of prostate cancer DU145, *Exp Mol Pathol* 88(1) 90-95.
- [55] P. Chavrier, J. M é n é trey, Toward a Structural Understanding of Arf Family:Effector Specificity, *Cell* 18 (2010) 1552-1558.
- [56] J.S. Bonifacio, The GGA proteins: adaptors on the move, *Nat Rev Mol Cell Biol* 5 (2004) 23 – 32.
- [57] A.K. Gillingham, S. Munro, The Small G Proteins of the Arf Family and Their Regulators, *Rev Cell Dev Biol* 23 (2007) 579 – 611.
- [58] J. R. Benjamina, P. Poona, J.D. Drysdale, X.M. Wang, R.A. Singer, G.C. Johnston, Dysregulated Arl1, a regulator of post-Golgi vesicle tethering, can inhibit endosomal transport and cell proliferation in yeast, *Mol Biol* 22(13) (2011) 2337-2347.
- [59] L. Lu, H. Horstmann, C. Ng, W. Hong, Regulation of Golgi structure and function by ARF-like protein 1 (Arl1), *J. Cell Sci* 114(2001) 4543 – 4555.
- [60] C.G. Burd, T.I. Strohlic, S.R. Gangi Setty, Arf-like GTPases: not so Arf-like after all, *Trends in Cell Biol* 14(12) (2004) 687-694.
- [61] L. Kjer-Nielsen, R.D. Teasdale, C. Van Vliet, P.A. Gleeson, A novel Golgi-localisation domain shared by a class of coiled-coil peripheral membrane proteins, *Curr. Biol* 9(1999) 385 – 388.
- [62] S. Munro, B.J. Nichols, The GRIP domain—a novel Golgi-targeting domain found in several coiled-coil proteins, *Curr. Biol* 9(1999) 377 – 380.
- [63] H. Van Valkenburgh, J.F. Shern, J.D. Sharer, X. Zhu, R.A. Kahn, ADP-ribosylation factors (ARFs) and ARF-like 1 (ARL1) have both specific and shared effectors: characterizing ARL1-binding proteins, *J Biol Chem* 276(2001) 22826 – 22837.
- [64] B. Panic, J.R. Whyte, S. Munro, The ARF-like GTPases Arl1p and Arl3p Act in a pathway that interacts with vesicle-tethering factors at the Golgi apparatus, *Curr Biol* 13(2003) 405 – 410.
- [65] J.W. Tamkun, R.A. Kahn, M. Kissinger, B. Brizuela, C. Rulka, M. P. Scott, J.A. Kennison, The arflike gene encodes an essential GTP-binding protein in *Drosophila*, *Proc Nat Acad Sci* 88(1991) 3120-3124.
- [66] F.J. Lee, C.F. Huang, W.L. Yu, L.M. Buu, C.Y. Lin, M.C. Huang, J. Moss, M. Vaughan, Characterization of an ADP-ribosylation factor-like 1 protein in *Saccharomyces cerevisiae*, *J Biol Chem* 272 (1997) 30998-31005.
- [67] K. Haglund, I. Dikic, Ubiquitylation and cell signaling, *EMBO* 24 (2005) 3353 – 3359.
- [68] N. Jura, E. Scotto-Lavino, A. Sobczyk, D. Bar-Sagi, Differential modification of Ras proteins by ubiquitination, *Mol Cell* 21(5) (2006) 679-687.
- [69] A. Lu, F. Tebar, B. Alvarez-Moya, C. López-Alcalá, M. Calvo, C. Enrich, et al, A clathrin-dependent pathway leads to KRas signaling on late endosomes en route to lysosomes, *J Cell Biol* 184(6) (2009) 863-879.
- [70] L. Xu, V. Lubkov, L.J. Taylor, D. Bar-Sagi, Feedback Regulation of Ras Signaling by Rabex-5-Mediated Ubiquitination, *Curr Biol* 20(2010) 1372-1377.
- [71] S.E. Kim, J.Y. Yoon, W.J. Jeong, S.H. Jeon, Y. Park, J.B. Yoon, et al, H-ras is degraded by Wnt/beta-catenin signaling via beta-TrCP-mediated polyubiquitylation, *J Cell Sci* 122 (2009) 842-848.

- [72] A.T. Sasaki, A. Carracedo, J.W. Locasale, K. Anastasiou, D. Takeuchi, E.R. Kahoud, et al. Ubiquitination of K-ras enhances activation and facilitates binding to select downstream effectors, *Sci Signal* (2011) 4-13.
- [73] A. Doye, A. Mettouchi, G. Bossis, R. Clement, C. Buisson-Touati, G. Flatau, L. Gagnoux, M. Piechaczyk, P. Boquet, E. Lemichez, CNF1 exploits the ubiquitin-proteasome machinery to restrict Rho GTPase activation for bacterial host cell invasion, *Cell* 111(4) (2002) 553-564.
- [74] M. Barrios-Rodiles, K.R. Brown, B. Ozdamar, R. Bose, Z. Liu, R.S. Donovan, F. Shinjo, Y. Liu, J. Dembowy, I.W. Taylor, V. Luga, N. Przulj, M. Robinson, H. Suzuki, Y. Hayashizaki, I. Jurisica, J.L. Wrana, High-throughput mapping of a dynamic signaling network in mammalian cells, *Science* 307(5715) (2005) 1621-1625.
- [75] T. Liedtke, J.C. Schwamborn, U. Schroer, S. Thanos, Elongation of axons during regeneration involves retinal crystallin beta b2 (crybb2), *Mol Cell Proteomics* 6(5) (2007) 895-907.
- [76] H. Kawabe, A. Neeb, K. Dimova, S.M. Young, M. Takeda, S. Katsurabayashi, M. Mitkovski, O.A. Malakhov, D.E. Zhang, M. Umikawa, K. Kariya, S. Goebbels, K. Armin Nave, C. Rosenmund, O. Jahn, J. Rhee, N. Brose, Regulation of Rap2A by the Ubiquitin Ligase Nedd4-1 Controls Neurite Development, *Cell Neuron* 65(2010)358-472.
- [77] A. Sakane, S. Hatakeyama, T. Sasaki, Involvement of Rabring7 in EGF receptor degradation as an E3 ligase, *Biochem Biophys Res Commun* 357 (2007) 1058-1064.
- [78] S.J. Plowman, R.L. Berry, S.A. Bader, F. Luo, M.J. Arends, D.J. Harrison, M.L. Hooper, C.E. Patek, K-ras 4A and 4B are Co-expressed Widely in Human Tissues and Their Ratio is Altered in Sporadic Colorectal Cancer, *J Exp Clin Cancer Res* 25(2) (2006) 259-267.
- [79] Fritz, G. & Kaina, B. Ras-related GTPase RhoB represses NF- $\kappa$ B signaling. *J. Biol. Chem.* (2001) 276, 3115 – 3122
- [80] Zhu H, Kavsak P, Abdollah S, et al. A SMAD ubiquitin ligase targets the BMP pathway and affects embryonic pattern formation[J]. *Nature*, 1999, 400(6745):687-693.
- [81] Narimatsu M, Bose R, Pye M, et al. Regulation of planar cell polarity by Smurf ubiquitin ligases[J]. *Cell*, 2009, 137(2):295-307.
- [82] Tajima Y, Goto K, Yoshida M, et al. Chromosomal region maintenance 1 (CRM1)-dependent nuclear export of Smad ubiquitin regulatory factor 1 (Smurf1) is essential for negative regulation of transforming growth factor-beta signaling by Smad7[J]. *J Biol Chem*, 2003, 278(12):10716-10721.
- [83] Wang H R, Zhang Y, Ozdamar B, et al. Regulation of cell polarity and protrusion formation by targeting RhoA for degradation[J]. *Science*, 2003, 302(5651):1775-1779.
- [84] Yamashita M, Ying S X, Zhang G M, et al. Ubiquitin ligase Smurf1 controls osteoblast activity and bone homeostasis by targeting MEKK2 for degradation[J]. *Cell*, 2005, 121(1):101-113.
- [85] Andrews PS, Schneider S, Yang E, Michaels M, Chen H, Tang J, Emkey R. Identification of substrates of SMURF1 ubiquitin ligase activity utilizing protein microarrays

Degree papers are in the “[Xiamen University Electronic Theses and Dissertations Database](#)”. Full texts are available in the following ways:

1. If your library is a CALIS member libraries, please log on <http://etd.calis.edu.cn/> and submit requests online, or consult the interlibrary loan department in your library.
2. For users of non-CALIS member libraries, please mail to [etd@xmu.edu.cn](mailto:etd@xmu.edu.cn) for delivery details.

厦门大学博硕士论文摘要库